Analyzing Cytotoxicity in a 2D and 3D Colorectal Cancer Model using an Automated High Content Imaging System

Schaefer, W.1; Hedemann, N.2; Willms, A.1; Werdelmann, B.1; Stoehr, M.1; Schulze, K.1; Sebens, S.3; Geisen, R.1 & Pirsch, M.1

¹SYNENTEC GmbH, Elmshorn, Germany; ²Department of Gynaecology and Obstetrics, CAU + UKSH, Kiel, Germany; ³Institute for Experimental Cancer Research, CAU + UKSH, Kiel, Germany

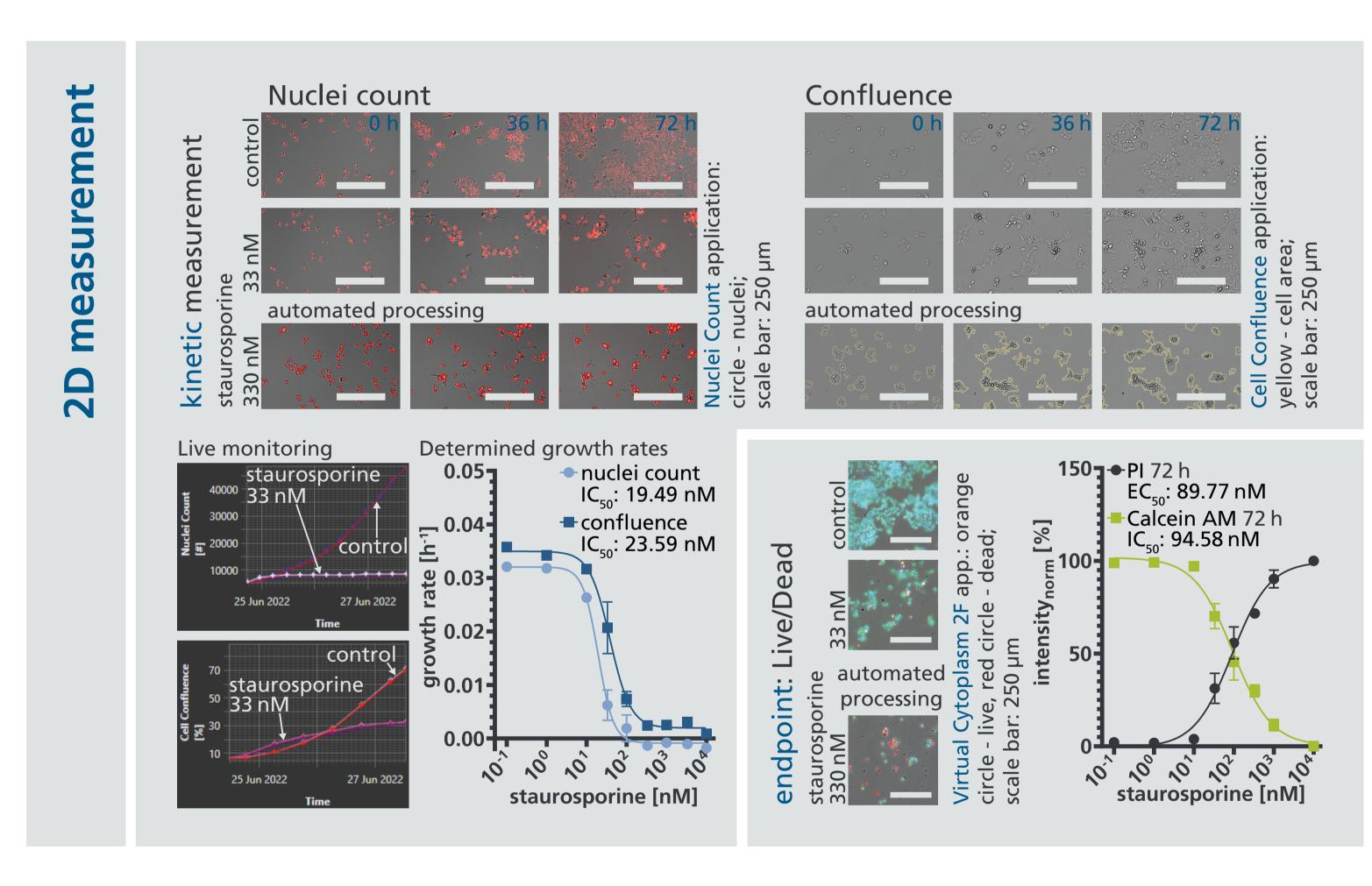
Introduction Method Seeding colorectal cancer cells HCT116 in 96 well plate Adhesion **Spheroid** formation for 24 h over 4 days Treatment for 72 h Adding CellTox SPY650-DNA to assay to treatment treatment ! _ _ Imaging every 6 h over 72 h for 10 min Live/Dead assay Imaging after every 2 h 72 h over 72 h for 2 min **Automated** high-throughput or manuell imaging

3D and 2D analysis with our automation system:

- The system enables convenient and timesaving high-throughput imaging
- Cytotoxic effects can be monitored directly over time
- YT-SOFTWARE® analyzes many different parameters
- Time charts and heat maps provide a quick result overview
- Various assays can be performed
- 3D can be easily compared to 2D

An important tool in the field of cancer research and drug discovery is cell culture. Conventionally, two-dimensional (2D) cell culture models have been used, but in the last years, three-dimensional (3D) cell culture models like spheroids have gained more interest. 3D models better mimic the physiology of tissues and tumors, but the generation, application and evaluation of assays is usually more difficult. Therefore, either simple one-parameter endpoint assays using plate readers are used as a read-out for high-throughput screening. Or confocal or time-lapse microscopes are used for multi-parameter high content analysis.

> Here, we used SYNENTEC's automation system to analyze effects of the drug staurosporine in 2D and 3D cell models of colorectal cancer cells (HCT116) over time.



3D measurement

